

# Characterization of Photochemical Processes for H<sub>2</sub> Production by CdS Nanorod–[FeFe] Hydrogenase Complexes

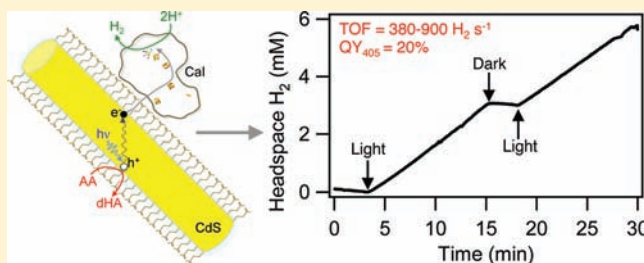
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**S** Supporting Information

**ABSTRACT:** We have developed complexes of CdS nanorods capped with 3-mercaptopropionic acid (MPA) and *Clostridium acetobutylicum* [FeFe]-hydrogenase I (CaI) that photocatalyze reduction of H<sup>+</sup> to H<sub>2</sub> at a CaI turnover frequency of 380–900 s<sup>-1</sup> and photon conversion efficiencies of up to 20% under illumination at 405 nm. In this paper, we focus on the compositional and mechanistic aspects of CdS:CaI complexes that control the photochemical conversion of solar energy into H<sub>2</sub>. Self-assembly of CdS with CaI was driven by electrostatics, demonstrated as the inhibition of ferredoxin-mediated H<sub>2</sub> evolution by CaI. Production of H<sub>2</sub> by CdS:CaI was observed only under illumination and only in the presence of a sacrificial donor. We explored the effects of the CdS:CaI molar ratio, sacrificial donor concentration, and light intensity on photocatalytic H<sub>2</sub> production, which were interpreted on the basis of contributions to electron transfer, hole transfer, or rate of photon absorption, respectively. Each parameter was found to have pronounced effects on the CdS:CaI photocatalytic activity. Specifically, we found that under 405 nm light at an intensity equivalent to total AM 1.5 solar flux, H<sub>2</sub> production was limited by the rate of photon absorption (~1 ms<sup>-1</sup>) and not by the turnover of CaI. Complexes were capable of H<sub>2</sub> production for up to 4 h with a total turnover number of 10<sup>6</sup> before photocatalytic activity was lost. This loss correlated with inactivation of CaI, resulting from the photo-oxidation of the CdS capping ligand MPA.



## INTRODUCTION

The capture and conversion of solar energy to couple water oxidation to carbon fixation or hydrogen (H<sub>2</sub>) production is an active area for developing renewable fuel technologies.<sup>1–6</sup> Studies of photosynthetic H<sub>2</sub> production,<sup>7–12</sup> first demonstrated in 1942,<sup>13</sup> have been models for inspiring the development of synthetic photochemical systems that are both more efficient and robust.<sup>14</sup> At the intersection of these two approaches, biohybrids integrate biological and synthetic molecules to engineer new photocatalytic assemblies for solar conversion. For example, photosystem I (PSI) has been assembled with inorganic H<sub>2</sub> production catalysts such as Pt nanoparticles<sup>15–17</sup> and transition metal compounds<sup>18</sup> or linked directly to H<sub>2</sub> activating enzymes, hydrogenases (H<sub>2</sub>ases).<sup>19,20</sup> Complementary approaches have coupled H<sub>2</sub>ases to particulate semiconductor materials<sup>21–23</sup> or onto conductive electrodes in photoelectrochemical devices.<sup>24,25</sup> In addition to these, there are many examples of completely synthetic photochemical systems,<sup>26–29</sup> including designs that couple semiconductors with inorganic surface catalysts.<sup>30–33</sup>

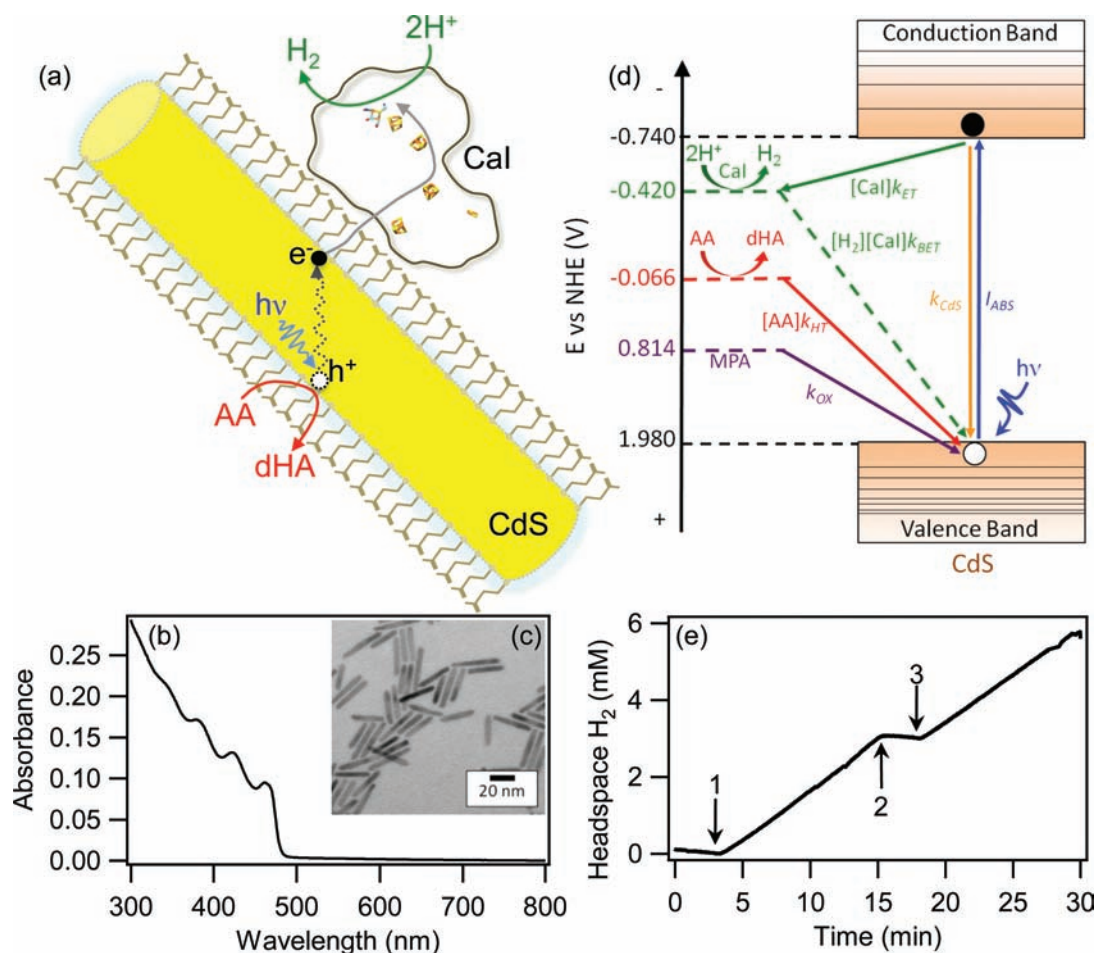
As H<sub>2</sub> activation catalysts, H<sub>2</sub>ases exhibit some ideal characteristics including low activation energies, variation in bias, and a wide range of O<sub>2</sub> sensitivities, while utilizing organometallic catalytic sites composed of earth abundant metals.<sup>34–40</sup> Recently, single-molecule resolution electrochemistry experiments estimated the turnover frequency (TOF) of

H<sub>2</sub> production by the [FeFe]-H<sub>2</sub>ase from *Clostridium acetobutylicum* (CaI) at 21 000 s<sup>-1</sup> at an overpotential of 150 mV.<sup>41</sup> This value for H<sub>2</sub> production TOF is among the highest for any H<sub>2</sub>ase,<sup>42,43</sup> underscoring the potential of CaI as a model catalyst for developing biohybrids to investigate functional requirements for photocatalytic H<sub>2</sub> production.

We have shown that CaI forms catalytically active complexes with a variety of nanoparticles and functions as an electrocatalyst on nanostructured electrodes.<sup>44,45</sup> In particular, CaI forms photocatalytic complexes with 3-mercaptopropionic acid (MPA) capped CdTe nanocrystals.<sup>21</sup> These complexes displayed relatively short time scales of H<sub>2</sub> production and tended to decompose under illumination.<sup>21</sup> In the work described here, we have employed colloidal CdS nanorods to achieve greater photostability combined with a larger absorption cross-section and surface area. This material possesses excellent light-harvesting characteristics (molar absorptivities ~10<sup>7</sup> M<sup>-1</sup> cm<sup>-1</sup> at 405 nm, as described later in the text), and both bulk and nanoscale CdS have appropriate band-edge energies for water oxidation and proton reduction.<sup>31,46–50</sup> A number of photocatalytic systems derived from colloidal CdS nanorods attached to metallic catalysts have been reported to produce H<sub>2</sub> under solar irradiation.<sup>30,31,49,51</sup>

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**Figure 1.** Properties of CdS nanorods and CdS:CaI complexes. (a) The proposed scheme for photocatalytic H<sub>2</sub> production by CdS:CaI complexes. Electron-transfer [FeS]-clusters and the catalytic H-cluster are shown in yellow (sulfur) and orange (iron). CdS and CaI are drawn to scale, while MPA molecules are enlarged  $\sim 5\times$ . (b) UV-vis absorption spectrum of CdS nanorods in water. (c) TEM of typical CdS nanorods, average size  $30 \times 4.5$  nm based on measurement of at least 200 nanorods. (d) Energy level diagram showing processes in photoexcited CdS that are relevant to H<sub>2</sub> production. (Abbreviations: AA, ascorbic acid; dHA, dehydroascorbate; CaI, [FeFe]-hydrogenase;  $E_g$ , band gap energy;  $k_{ET}$ , rate constant for ET from CdS to CaI;  $k_{HT}$ , rate constant for HT from CdS to AA; MPA, mercaptopropionic acid;  $k_{OX}$ , rate of photo-oxidation of surface-bound MPA ligands;  $k_{BET}$ , rate constant for ET from CaI to CdS (H<sub>2</sub> uptake);  $I_{ABS}$ , flux of absorbed photons;  $k_{Cds}$ , rate of excited state decay in CdS, including both radiative and nonradiative pathways (e.g., electron-hole recombination, carrier trapping). Potentials are shown vs NHE (pH 7, 1 atm H<sub>2</sub>). (e) Photocatalytic H<sub>2</sub> production by CdS:CaI complexes in 50 mM Tris-HCl, 275 nM CdS, 360 nM CaI, and 100 mM AA illuminated with white light at  $10\,000 \mu E m^{-2} s^{-1}$ . Arrows show changes in illumination: (1) light on, (2) light off, (3) light on.

Furthermore, CdS nanocrystals can be functionalized with mercapto-carboxylic acids as capping ligands.<sup>31,52</sup> Such ligands promote adsorption of CaI on both quantum dots<sup>21</sup> and electrodes<sup>41,53</sup> and stabilize inorganic nanomaterials in aqueous buffers that are required for enzyme function.

Here we characterize the parameters controlling the photocatalytic H<sub>2</sub> production rates in biohybrid assemblies composed of [FeFe]-H<sub>2</sub>ase CaI and MPA-capped CdS. The thermodynamics of self-assembly and the resulting orientations were investigated using kinetic analysis of CdS binding to CaI in competition with ferredoxin (Fd). We characterized the effect of CaI coverage in the context of CdS photon absorption and CaI catalytic flux. We also investigated the effects of hole-transfer and light intensity and found the H<sub>2</sub> production to be photon-limited under conditions approximating AM 1.5 solar flux. The quantum yield (QY) of the photocatalytic H<sub>2</sub> production was found to be  $\sim 20\%$ . New insights on the instability observed for biohybrids under illumination, first observed for MPA-CdTe:CaI complexes,<sup>21</sup> are also provided. These analyses demonstrate how the contributions of individual

processes govern photocatalytic activity and offer guidance in terms of the molecular design of future artificial photosynthetic systems.

## MATERIALS AND METHODS

**[FeFe]-hydrogenase Expression and Purification.** The StrepII-tagged [FeFe]-H<sub>2</sub>ase from *C. acetobutylicum* was expressed and purified from *Escherichia coli* as previously described.<sup>54</sup> Specific activities of CaI preparations were measured as the H<sub>2</sub> evolved from sodium dithionite (Riedel-de Haen) reduced methyl viologen (Sigma) (MV, 5 mM).<sup>55</sup> Purified CaI had specific activities of  $\sim 1300 \mu mol H_2 mg^{-1} min^{-1}$ , with protein concentrations determined by the Bradford assay using hemoglobin as the standard.<sup>56</sup>

**Ferredoxin Expression and Purification.** The mature form of the Fd protein PetF of *Chlamydomonas reinhardtii* was cloned and expressed in *E. coli* as described in the Supporting Information.

**Synthesis of CdS Nanorods.** The synthesis was adapted from previously published procedures (Supporting Information).<sup>57,58</sup> Synthesis and processing were conducted under an inert argon atmosphere at  $\sim 620$  Torr (atmospheric pressure in Boulder, CO). The resulting solutions consisted of nanorods with average diameter of

4.5 nm and average length of 30 nm. Molar absorptivity ( $\epsilon$ ) for CdS nanorods was determined by correlating absorption spectra with Cd<sup>2+</sup> concentrations determined by elemental analysis (ICP-OES) of acid-digested samples. The value of  $\epsilon$  was estimated to be 1710 M<sup>-1</sup> cm<sup>-1</sup> per Cd<sup>2+</sup> at 350 nm. The number of Cd<sup>2+</sup> per nanorod was estimated from average length and diameter measurements determined from TEM images.

**Ligand Exchange on CdS Surface.** The ligand exchange procedure was adapted from Amirav et al.<sup>31</sup> First, 1.27 mmol of 3-mercaptopropionic acid (3-MPA, Sigma Aldrich  $\geq 99\%$ ) was dissolved in 20 mL of methanol. The solution pH was increased to 11 with tetramethylammonium hydroxidepentahydrate salt (Sigma Aldrich). A sample of nanocrystals was precipitated from toluene solution using methanol. The precipitated nanocrystals were then mixed with the MPA/methanol solution until the mixture was no longer cloudy. The water-soluble nanocrystals were precipitated with toluene. The resulting particles were dried under vacuum and redispersed in Tris buffer, pH 7.

**Transmission Electron Microscopy.** TEM samples were prepared by drop casting on carbon film, 300 mesh copper grids from Electron Microscopy Sciences. Images were taken on a 100 KV Phillips CM100 transmission electron microscope equipped with a bottom-mounted 4 megapixel AMT v600 digital camera. Lengths and diameters were determined from an average of at least 200 particles.

**Determination of Reduction Potential of CdS Nanorods.** Mixtures of 83 nM CdS nanorods and 167 nM MV (Sigma) were combined under an anaerobic Ar atmosphere in buffers of varying pH (2, 2.5, 3, 3.4, 4, and 4.5).<sup>59,60</sup> Samples were illuminated for 10 min with 405 nm LED light at 800  $\mu\text{E m}^{-2} \text{s}^{-1}$  intensity. Absorbance spectra were collected on a Beckman DU800 and the concentration of reduced MV was determined by absorbance at 606 nm ( $A_{606}$ ) (Supporting Information). The potential of the conduction band electrons in photoexcited CdS at standard reaction conditions was determined by extrapolation to pH 7 using the Nernst equation.

**Assembly of CdS:CaI Complexes.** Mixtures of CdS nanorod and CaI solutions for all experiments were prepared in buffer (50 mM Tris-HCl, 5 mM NaCl, 5% glycerol, pH 7), under an anaerobic atmosphere of 4% H<sub>2</sub>/96% N<sub>2</sub>, and then sparged with 100% Ar. The CdS concentration was determined from absorbance at 350 nm ( $\epsilon = 9 \times 10^6 \text{ M}^{-1} \text{ cm}^{-1}$ ). Samples were stirred for at least 10 min to ensure complete assembly prior to all experiments (see Figure S2, Supporting Information).

**Ferredoxin–CdS Competition Kinetics.** Mixtures of CdS nanorods and CaI in molar ratios of 0:1, 2:1, 5:1, and 10:1 CdS:CaI were prepared under an anaerobic atmosphere of 4% H<sub>2</sub>/96% N<sub>2</sub> and then sparged with 100% Ar, with a fixed CaI concentration of 28 nM for all samples. Stock Fd solutions were reduced with 5 mM sodium dithionite and added to CdS:CaI mixtures to achieve the desired Fd concentration (0, 15, 30, 60, 150, 300  $\mu\text{M}$ ). The headspace H<sub>2</sub> was measured after 6 min by injection into an Agilent 7890A gas chromatograph fitted with a 5 Å molecular sieve column (Supelco). A constant low level of background H<sub>2</sub> production from reduction of CaI by 5 mM sodium dithionite was observed in all reactions and was subtracted from the sample containing Fd.

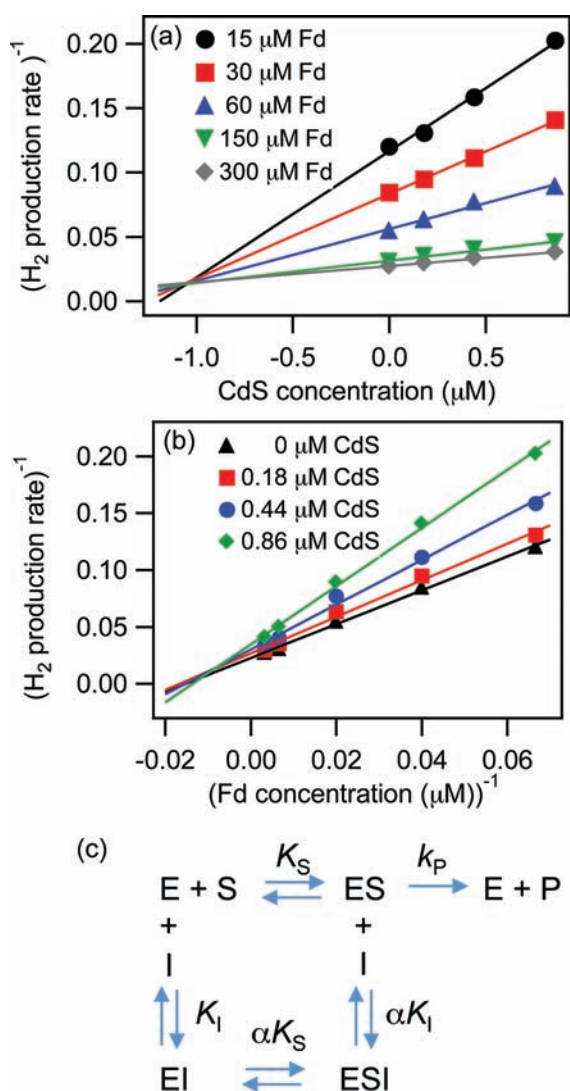
**Light-Driven H<sub>2</sub> Production.** Samples of CdS:CaI were prepared as above. Solutions contained 100 mM ascorbic acid (AA, Fluka) unless otherwise noted. Gas analysis of sample headspace confirmed H<sub>2</sub> removal. Samples were illuminated for 10 min with an Ocean Optics 405 nm LED light source (750 mW, 500 mA) at 800  $\mu\text{E m}^{-2} \text{s}^{-1}$  (light meter LI-250, LI-COR Biosciences) unless otherwise noted. The photon flux of AM 1.5 photons with sufficient energy to excite CdS nanorods ( $E_g = 464 \text{ nm}$ ) was calculated to be 450  $\mu\text{E m}^{-2} \text{s}^{-1}$ . Samples were incubated in 1.5 mL serum vials with an illuminated surface area of 5 mm<sup>2</sup> unless otherwise noted. Light intensities were confirmed before each illumination experiment. Headspace H<sub>2</sub> was measured at 10 min by GC injection or in real-time by continuous measurement by an Omnistar capillary mass spectrometer (Pfeiffer). Turnover frequencies (TOF) for photochemical H<sub>2</sub> production are expressed in units of mol H<sub>2</sub> (mol CaI)<sup>-1</sup> s<sup>-1</sup>.

**Quantum Yield Measurements.** Mixtures of CdS nanorods and CaI in molar ratios of 1:0.67 CdS:CaI (14 nM CdS, 9 nM CaI) were prepared as above in buffer with 100 mM AA. Samples were illuminated in a quartz cuvette with a 3 × 5 mm window with a 1 mW 405 nm laser pointer (Laserglow Technologies) with a spot size of 6 mm<sup>2</sup>. Light intensity passing through the sample was measured using a Newport model 1918-R power meter with an 818-ST-UV UV-enhanced silicon photodetector. The difference between light intensity transmitted through a control cuvette containing 50 mM Tris-HCl buffer (pH 7) and through the sample cuvette corresponded to absorbed intensity. The amount of H<sub>2</sub> produced was measured after 10 min of illumination by GC. The quantum yield was defined as QY = (mol H<sub>2</sub> produced/mol photons absorbed) × (2 mol photons/mol H<sub>2</sub>).

## RESULTS AND DISCUSSION

**Assembly and Orientation of Photocatalytically Active CdS:CaI Complexes.** Figure 1a shows a structural model of the complex formed between CaI and MPA-capped CdS nanorods (referred to here as CdS), with a predicted conformation based on the surface charges of the two components. The MPA bonds to the CdS surface via the thiol group, with the solvent-exposed carboxylate group imparting a negative surface charge at pH 7. As previously shown, MPA-capped nanoparticles formed complex with CaI via electrostatic interactions and were predicted to orient the positively charged enzyme surface near the distal [4Fe–4S] cluster toward the MPA carboxylate group.<sup>21,61</sup> In a similar fashion, the CdS used here was expected to bind to this site on CaI, which is also the site for ferredoxin (Fd) interaction during ET in vivo.<sup>55,62</sup> When CdS and CaI were mixed, complexes self-assembled and reached full photocatalytic activity within 10 min (Figure S2, Supporting Information). The H<sub>2</sub> production rates were linear under illumination with white light in the presence of 100 mM AA, and no significant H<sub>2</sub> production was seen when the illumination is turned off (Figure 1e) or when any of the components (CdS, AA, or H<sub>2</sub>ase) were absent.

To confirm the predicted conformation model of the CdS:CaI binding complex, the effect of CdS on Fd-driven H<sub>2</sub> production by CaI was tested (Figure 2). The analysis of the inhibition kinetics by Dixon plot<sup>63,64</sup> (Figure 2a) or double-reciprocal plot<sup>65</sup> (Figure 2b) was consistent with a linear mixed-type inhibition, as described by the equilibrium model shown in Figure 2c. The values for  $K_i$ , the dissociation constant for inhibitor (CdS) and CaI binding, and  $\alpha$ , the factor by which  $K_S$  changes in the presence of CdS, are shown in Table 1. A key feature of mixed inhibition is the ability of the EI complex (CdS:CaI) to bind the substrate, reduced Fd, in a nonactive complex, ESL. Thus, as CdS concentration increases, the soluble Fd pool is effectively decreased, giving rise to the measured value of  $\alpha$ . For CdS:CaI, this behavior is likely explained by binding of the Fd to the CdS nanorod. Although the Fd surface that binds to CaI is negatively charged, the opposing face is more positively charged, forming a dipole.<sup>66</sup> Thus, Fd could bind to the negatively charged MPA–CdS via an electrostatic interaction, as has been shown in the case of MPA-modified Au surfaces.<sup>67</sup> On the basis of the  $K_i$  value, binding of CdS to CaI is stronger than that of Fd, possibly due to a larger contact surface between CdS and CaI and stronger electrostatic interaction. On the other hand, the Fd–CaI ET complex by necessity is exchangeable to allow for multiple reduction–oxidation reactions between Fd and other donor/acceptor molecules in the cell. By creating a stable CaI–CdS interaction, ET is no longer diffusion limited, and turnover rates of CaI



**Figure 2.** Kinetics of CdS inhibition of Fd-mediated CaI H<sub>2</sub> evolution in the dark. The CaI concentration was fixed at 28 nM, and the CdS concentration was varied between 0 and 1.1 μM to achieve molar ratios of 0:1, 1.5:1, 3.8:1, and 7.5:1 CdS:CaI. Reduced Fd (in 2.5 mM sodium dithionite) was added to yield final concentrations of 15, 30, 60, 150, and 300 μM. The total production of H<sub>2</sub> was measured after 6 min in the dark. The measured  $K_M$  ( $K_S$  in the equation) of CaI for Fd in the absence of CdS was found to be 44 μM. (a) Dixon plot of the inverse of H<sub>2</sub> production TOF (mol<sup>-1</sup> H<sub>2</sub> mol CaI s) versus CdS concentration (μM). The measured  $K_I$  of CdS is ~1.4 μM (average of the values shown in Table 1). (b) A double-reciprocal plot of inverse H<sub>2</sub> production rate versus inverse Fd concentration. (c) Equilibrium model of linear mixed-type inhibition, where E is CaI, I is CdS, S is Fd, and P is H<sub>2</sub>.

**Table 1. Inhibition Constants for CdS on Fd-Mediated H<sub>2</sub> Evolution by CaI**

analysis method <sup>a</sup>	kinetic constants
Dixon plot, [CdS] vs (TOF) <sup>-1</sup>	$K_I = 1.28 \pm 0.19 \mu\text{M}$ $\alpha = 2.77 \pm 0.58$
double reciprocal, [Fd] <sup>-1</sup> vs (TOF) <sup>-1</sup>	$K_I = 1.58 \pm 0.26 \mu\text{M}$ $\alpha = 2.31 \pm 0.38$

<sup>a</sup>TOF = mol H<sub>2</sub> (mol CaI)<sup>-1</sup> s<sup>-1</sup>.

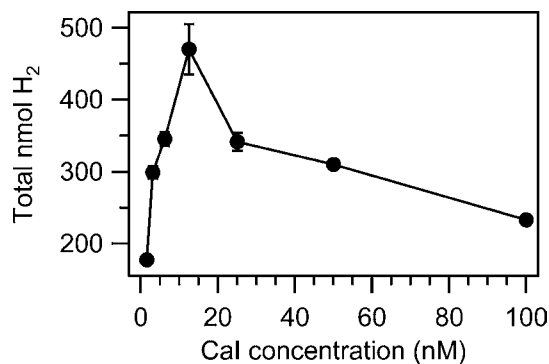
might approach those measured using electrochemical techniques.

**Determination of the CdS Band-Edge Reduction and Oxidation Potentials.** The band structure of CdS and the formal potentials of the charge carrier acceptors are shown in Figure 1d. The conduction band (photoexcited electron) potential in CdS was measured in solution by photoreduction of the redox-active dye methyl viologen (MV<sup>2+</sup>), for a range of pH values (Figure S1, Supporting Information).<sup>59,60</sup> The valence band (hole) potential was calculated by subtracting the CdS optical band gap energy (464 nm, 2.9 eV, Figure 1b) from the measured potential of the conduction band. The CaI operating potential is estimated by the Nernst equation of the H<sup>+</sup>/H<sub>2</sub> redox couple (pH 7, 1 atm H<sub>2</sub>).<sup>55</sup> The overpotential of the CdS conduction band is therefore ~320 mV (vs NHE). Also shown is the electrochemical potential of the ascorbic acid (AA)–dehydroascorbate (dHA) redox couple,<sup>68</sup> which is used here as the sacrificial electron donor to regenerate the CdS ground state.

#### Effect of the CaI:CdS Ratio on Photocatalytic Rates.

The scheme shown in Figure 1d illustrates the photochemical processes that contribute to H<sub>2</sub> production activity of CdS:CaI complexes. Photon absorption by CdS ( $I_{\text{ABS}}$ ) results in the generation of electron–hole pairs. These are coupled to catalysis through the transfer of conduction band electrons to CaI at a rate that is proportional to the product of [CaI] $k_{\text{ET}}$  (referred to as “ET”), and hole scavenging by oxidation of AA at a rate proportional to [AA] $k_{\text{HT}}$  (referred to as “HT”). These external processes compete with oxidation of the MPA ligands ( $k_{\text{OX}}$ )<sup>31,69</sup> and internal recombination pathways in CdS ( $k_{\text{CdS}}$ ). The individual contributions of  $I_{\text{ABS}}$ , ET, and HT can be manipulated by changes in light intensities, the stoichiometric distribution of molecular complexes, and AA concentration, respectively. Adjusting the relative contribution of each process allows for a more complete understanding of how photon energy is allocated and how each process affects the overall conversion efficiency.

The first parameter tested was the ET rate, which was varied by changing the molar ratio of CaI molecules per CdS nanorod (rate of ET  $\propto$  [CaI] $k_{\text{ET}}$ ) and measuring the photocatalytic H<sub>2</sub> production rates (Figure 3). The light intensity and the concentrations of AA and CdS were fixed to ensure that the rates of photon absorption ( $I_{\text{ABS}}$ ), HT ( $\propto$  [AA] $k_{\text{HT}}$ ), and the



**Figure 3.** Effect of complex molar ratio of photocatalytic H<sub>2</sub> production by CdS:CaI. Mixtures prepared with a fixed CdS concentration of 19 nM and CaI concentrations between 1.58 and 100 nM to give molar ratios of 1:5, 1:2.5, 1:1.33, 1:0.67, 1:0.33, 1:0.167, and 1:0.083 CdS:CaI solutions. Samples were illuminated for 10 min with a 405 nm LED at 800 μE m<sup>-2</sup> s<sup>-1</sup>.

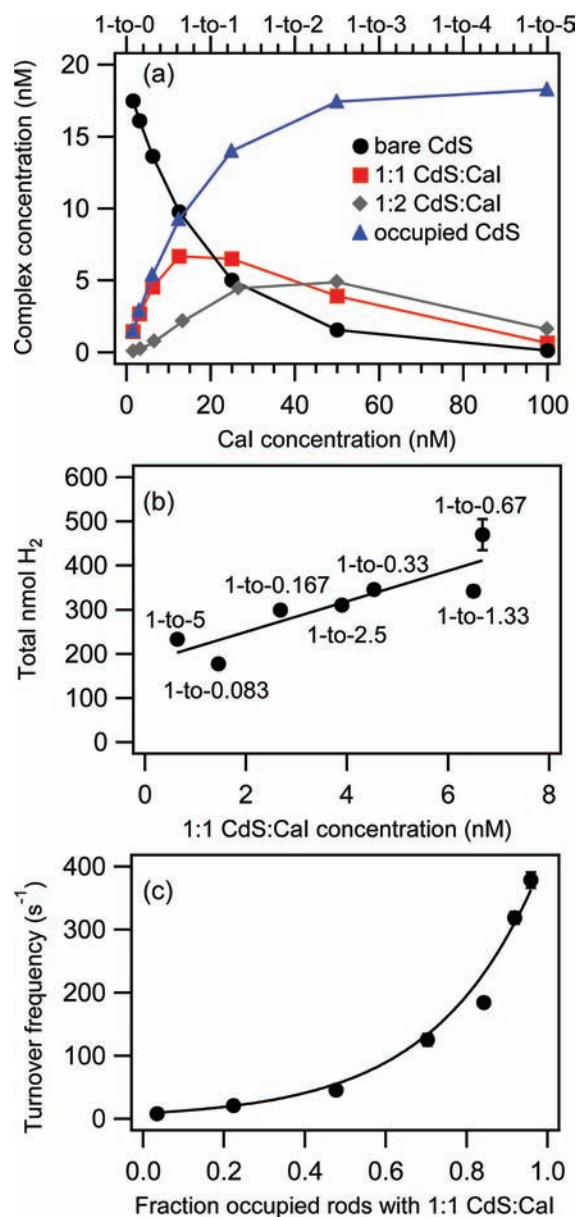
number of available photogenerated electrons were constant across all samples. A 405 nm light intensity of  $800 \mu\text{E m}^{-2} \text{s}^{-1}$  was used to approximate the flux AM 1.5 photons absorbed by CdS. We observed that  $\text{H}_2$  production rates initially increased with increasing CaI concentration up to 12.5 nM CaI (molar ratio 1:0.67 CdS:CaI) but then declined at higher CaI concentrations. We had expected that as CaI concentration increased the level of  $\text{H}_2$  production would also increase, since the fraction of the photoexcited electrons decaying by ET, rather than internal recombination in CdS, should increase with CaI concentration. We hypothesize that the decline in rate at high CaI coverage is due to back electron transfer (BET) arising from (i) CaI oxidation of  $\text{H}_2$  (Figure 1d,  $k_{\text{BET}}$ ) and/or (ii) competition for electrons among multiple CaI molecules. Both effects would reduce the  $\text{H}_2$  production TOF and total  $\text{H}_2$  levels in samples with high CaI:CdS ratios and multiple CaI molecules per CdS.

**Statistical Analysis of CdS:CaI Molar Ratio.** If the hypothesis above is valid, we would expect that complexes composed of a single CaI bound to a single CdS would exhibit peak  $\text{H}_2$  production efficiencies that decrease as multiple CaI molecules bind to each CdS. Thus, the amount of  $\text{H}_2$  generated by a solution of CdS:CaI complexes would scale with the relative concentration of the 1:1 fraction.

To test this model, a statistical analysis of the molecular compositions of each ratio was compared to the photocatalytic  $\text{H}_2$  production rates and TOF. Since the surface area of CdS nanorods ( $\sim 456 \text{ nm}^2$ ) is significantly larger ( $\sim 26$  times) than the binding surface of CaI ( $17.5 \text{ nm}^2$ ),<sup>61</sup> each nanorod should be able to accommodate several molecules of CaI (5–10) before steric effects limit additional binding. Thus, the probability of CaI binding to CdS was assumed to be independent of whether another CaI was present (up to the limit of 10 CaI molecules per CdS), and the fractional population of each ratio was modeled using a Poisson distribution.

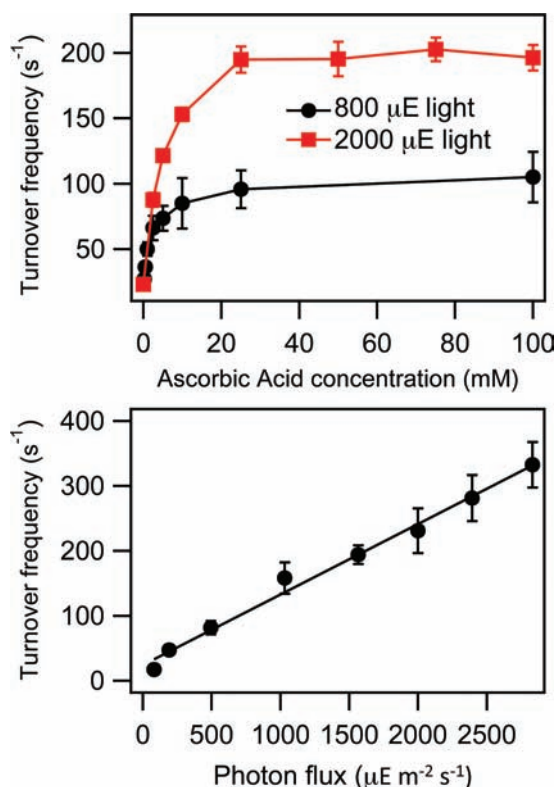
As shown in Figure 4a, the distribution of 1:1 and 2:1 CdS:CaI complexes (complete distributions for each molar ratio are listed in Table S1, Supporting Information) changes as a function of CaI concentration. Using the analysis performed in Figure 4a, the concentration of 1:1 complexes for each ratio was plotted versus the total  $\text{H}_2$  produced. A linear fit of the plot (Figure 4b) shows that as the fractional level of 1:1 increased, rates also increased. Furthermore, a plot of the 1:1 fraction (of all *occupied* nanorods) against the CaI TOF for each ratio predicts that a maximal TOF value of  $412 \text{ s}^{-1}$  (under 405 nm light at  $800 \mu\text{E m}^{-2} \text{s}^{-1}$ ) should be observed for a 100% solution of 1:1 complexes. Together the data presented in Figure 4 show that the 1:1 complexes, under these conditions, make the most efficient use of CaI to achieve the highest TOF values and  $\text{H}_2$  production rates.

**Contribution of AA and HT to CdS:CaI Photocatalytic Rates.** In order to investigate the effect of the contribution of HT, we examined the effect of AA concentration on photocatalytic  $\text{H}_2$  production by a CdS:CaI (1:0.67 CdS:CaI molar ratio) under two different intensities of 405 nm light (Figure 5a). In this case, the contribution of HT is changing, but the photon flux, complex concentrations, and ratio distributions are fixed. Under both light intensities, the TOF of the complexes increased to maximal values as the AA concentrations increased. At  $800 \mu\text{E m}^{-2} \text{s}^{-1}$  (black circles) the TOF for  $\text{H}_2$  production began to saturate at  $\sim 10 \text{ mM}$  AA and at  $\sim 25 \text{ mM}$  AA under illumination at  $2000 \mu\text{E m}^{-2} \text{s}^{-1}$  (red



**Figure 4.** Statistical analysis of photocatalytic  $\text{H}_2$  production by CdS:CaI complexes. (a) Concentrations of specific molecular complexes within each solution of CdS:CaI. Concentrations are calculated using the Poisson distribution to determine the fraction of 19 nM CdS with a specific CaI coverage for the molar ratios of 1:5 (100 nM CaI), 1:2.5 (50 nM CaI), 1:1.33 (25 nM CaI), 1:0.67 (12.5 nM CaI), 1:0.33 (6.25 nM CaI), 1:0.167 (3.13 nM CaI), and 1:0.083 (1.56 nM CaI) CdS:CaI. Bare CdS (black circles) has no CaI adsorbed, 1:1 CdS:CaI (red squares) has one CaI adsorbed per CdS, 2:1 CdS:CaI (gray diamonds) has 2 CaI adsorbed per CdS, and total occupied CdS (blue triangles) is the sum of all CdS with  $\geq 1$  CaI adsorbed per CdS. (b) total nanomoles of  $\text{H}_2$  produced under 10 min illumination with a 405 nm LED at  $800 \mu\text{E m}^{-2} \text{s}^{-1}$  vs the concentration of 1:1 CdS:CaI calculated from the Poisson distribution for each CaI concentration. The solid trace is a linear fit. (c) CaI TOF [ $\text{mol H}_2 (\text{mol CaI})^{-1} \text{s}^{-1}$ ] vs the fraction of occupied nanorods that are 1:1 complexes, calculated from the Poisson distribution. The line is an exponential fit to  $y = Ae^{bx}$  ( $A = 7.17$ ,  $b = 4.05$ ).

squares). Thus, a range of 10–25 mM AA was required to achieve maximal photocatalytic rates. Michaelis–Menten kinetic analysis of the data presented in Figure 5a showed that the AA interaction with CdS has an average  $k_{\text{cat}}/K_m$  of  $10^4$



**Figure 5.** Effects of hole scavenger and light intensity on photocatalytic H<sub>2</sub> production by CdS:CaI complexes. (a) Effect of AA concentration on TOF of H<sub>2</sub> production [mol H<sub>2</sub> (mol CaI)<sup>-1</sup> s<sup>-1</sup>] by a 1:0.67 mixture of CdS:CaI. The sample concentrations were fixed at 14 nM CdS and 9 nM CaI. Samples were illuminated for 10 min with 405 nm light at 800 (black circles) or 2000 (red squares) μE m<sup>-2</sup> s<sup>-1</sup>. (b) Effect of light intensity on TOF of H<sub>2</sub> production [mol H<sub>2</sub> (mol CaI)<sup>-1</sup> s<sup>-1</sup>] by 1:0.67 CdS:CaI solutions. The sample concentrations were fixed at 14 nM CdS and 9 nM CaI, with 100 mM AA as the hole-scavenger. Each sample was prepared in a cuvette with a 3 × 5 mm window and illuminated with a 405 nm light source for 10 min at the designated intensity. Also shown is a linear fit (slope = 0.108 μE m<sup>-2</sup>; y intercept = 24.5 s<sup>-1</sup>).

s<sup>-1</sup> M<sup>-1</sup> during H<sub>2</sub> production (Table S3, Supporting Information), which is ~10<sup>6</sup> below the upper limit for diffusion-limited processes.<sup>70,71</sup> Moreover, although ~2200 AA (0.2 nm<sup>2</sup>) can bind per CdS, it must actively exchange during turnover to maintain charge balance in CdS. Binding constants of AA for cysteine capped CdS quantum dots have been measured at 1 mM,<sup>72</sup> consistent with the 1–5 mM *K<sub>m</sub>* values we measured here (Figure 5a and Table S3, Supporting Information). Thus, AA is required at greater than millimolar levels to maintain the fast TOF of the CdS:CaI complexes during photochemical H<sub>2</sub> production due to the weak, diffusion-limited reaction kinetics with the CdS.

**Quantum Yield of H<sub>2</sub> Production.** The QY of photocatalytic H<sub>2</sub> generation by the 1:0.67 CdS:CaI solution was measured using a 405 nm laser (Table S2, Supporting Information). This laser light source produces a collimated beam with a photon flux of 540 μE m<sup>-2</sup> s<sup>-1</sup>. Measurement of photons absorbed (~16% of total) and H<sub>2</sub> produced showed that the QY of CdS:CaI was 20.4 ± 1.9% based on two photons (one photon = one electron) per H<sub>2</sub>. This compares favorably to the photosynthetic efficiencies of microalgae.<sup>14,73</sup> It should be noted that this QY value is an average value for the distribution of molecular complexes present in the 1:0.67 ratio.

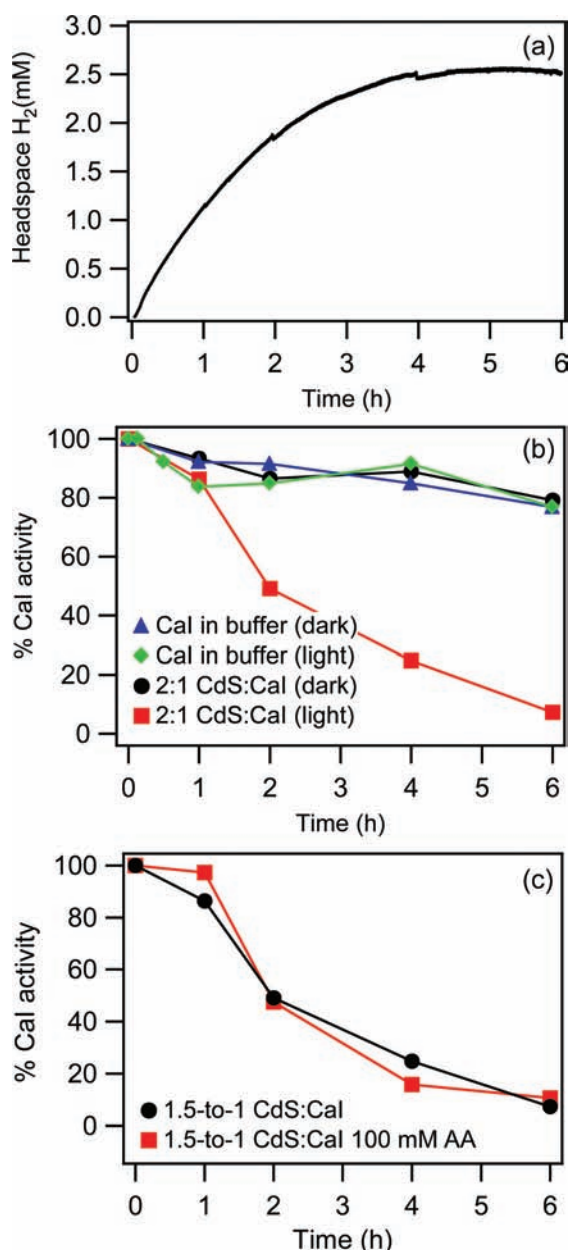
This will include a high fraction (~51%) of bare CdS, which is photocatalytically silent, and some with multiple (>1) CaI molecule coverage, which contribute to a decrease in H<sub>2</sub> production. Thus, it is likely that QY will vary with CdS:CaI molar ratio and that a solution consisting entirely of 1:1 coverage CdS:CaI would yield a higher QY value.

**Effect of Light Intensity on CdS:CaI Photocatalytic Rates.** A critical question in the design of biohybrid photocatalytic systems is whether the overall yield is limited by light absorption or the rate of catalysis (TOF). Recent measurements of CaI TOF on MPA–Au electrodes<sup>41</sup> can be used to estimate that for an overpotential of 320 mV, as is the case with CdS nanorods, the expected TOF would be ~4000 s<sup>-1</sup>. Since this value was not observed under the illumination conditions used here, H<sub>2</sub> production might be limited by the availability of photogenerated electrons. If this were the case, the H<sub>2</sub> production levels would be expected to increase with higher light intensities as the rate of the ET process approaches the maximum TOF of CaI.

Figure 5b shows that the H<sub>2</sub> generation rate scales linearly with light intensity (for the CdS:CaI ratio of 1:0.67). This indicates that the QY of H<sub>2</sub> production is constant over the intensity range examined and suggests that the rate at which the enzyme is able to convert the photoexcited electrons into H<sub>2</sub> is higher than the rate at which the electrons are supplied. At a photon flux of 540 μE m<sup>-2</sup> s<sup>-1</sup>, we estimate that each nanorod is excited on average 1000 times s<sup>-1</sup> (*e* = 6.5 × 10<sup>6</sup> M<sup>-1</sup> cm<sup>-1</sup>). At a H<sub>2</sub> production QY of 20%, this corresponds to an enzyme TOF of 100 s<sup>-1</sup>.

Using the linear fit of TOF vs light intensity in Figure 5b, the light intensity value for which electron flow (excitation rate × quantum efficiency of ET) will match CaI turnover for a 100% solution of 1:1 complexes can be estimated. Assuming that the response remains linear, the CaI TOF value of 4000 s<sup>-1</sup> will be reached at 405 nm light intensity of ~11 000 μE m<sup>-2</sup> s<sup>-1</sup>. This is ~25-fold higher than the integrated flux of AM 1.5 photons that can be used by CdS (450 μE m<sup>-2</sup> s<sup>-1</sup>, with λ < 464 nm) but similar in magnitude to full-spectral AM 1.5 flux of ~7000 μE m<sup>-2</sup> s<sup>-1</sup>. Thus, H<sub>2</sub> production in CdS:CaI complexes is photon-limited due to the absorption range of CdS, and efficiencies under solar illumination can be improved by widening the range of light-absorption, so that catalytic rates can be matched at solar flux, and by improving electron transfer efficiencies.

**Long-Term Behavior of CdS:CaI Complexes.** The long-term photocatalytic H<sub>2</sub> production by CdS:CaI is shown in Figure 6a. The sample consisted of a 1:0.67 mixture of CdS:CaI in 100 mM AA that was illuminated continuously for 6 h. The UV–vis spectra of the reactions during the 6 h period showed that AA oxidation (monitored as changes in absorbance at 360 nm)<sup>74,75</sup> occurred only in the presence of CaI (Figure S4, part a vs d, Supporting Information). The peak rate of H<sub>2</sub> production occurred within the first 30 min of illumination, followed by a slow, continuous decline in the observed rate throughout the next several hours. As a result, after 4 h of continuous illumination, there was little or no further accumulation of H<sub>2</sub>. The total turnover number (TON) for CdS:CaI was approximately 1 million. We tested several hypotheses to explain the noticeable decline in H<sub>2</sub> production rate after 4 h. (i) Since reactions are sealed, elevated H<sub>2</sub> levels lead to increased H<sub>2</sub> partial pressure for H<sub>2</sub> oxidation by CaI.<sup>55</sup> However, replacement of the headspace gas with 100% Ar every 10 min during illumination did not alter the long-term decline



**Figure 6.** Effects of long-term illumination of 1:0.67 CdS:CaI solution. (a) Photocatalytic H<sub>2</sub> production by a 1:0.67 CdS:CaI solution, 14 nM CdS, 9 nM CaI, 100 mM AA, illuminated 6 h with 800  $\mu\text{E m}^{-2} \text{s}^{-1}$  of 405 nm LED light. (b) Effect of illumination on CaI activity in CdS:CaI complexes. A concentration of 9 nM CaI in 50 mM Tris-HCl stirred for the appropriate time alone in the dark (blue triangle) or illuminated with 800  $\mu\text{E m}^{-2} \text{s}^{-1}$  of 405 nm LED light (green diamonds); mixture of 14 nM CdS, 9 nM CaI in 50 mM Tris-HCl stirred in the dark (black circles) or illuminated with 800  $\mu\text{E m}^{-2} \text{s}^{-1}$  of 405 nm LED light (red squares). CaI activity in all samples was measured in 5 mM MV at the specified time. (c) Effect of 100 mM AA on CaI long-term activity. A mixture of 14 nM CdS and 9 nM CaI in 50 mM Tris-HCl stirred under illumination with 800  $\mu\text{E m}^{-2} \text{s}^{-1}$  of 405 nm LED light with (red squares) and without (black circles) 100 mM AA.

in steady-state H<sub>2</sub> production rates (Figure S4, Supporting Information). (ii) Although the oxidation of AA was evident during H<sub>2</sub> production (time points 0–4 h, Figure S4a, Supporting Information), based on the net H<sub>2</sub> produced the residual [AA] remained well above the level needed to support

maximal rates (20 mM at 800  $\mu\text{E m}^{-2} \text{s}^{-1}$ , Figure 5a) and was not the limiting component. (iii) We also ruled out precipitation and degradation of the CdS nanorods. There was no evidence of significant changes in the UV–vis absorbance profile or in the dimensions based on TEM images (Figures S4 and S5, Supporting Information). Thus, none of these factors could account for the decline in photocatalytic rates shown in Figure 6a.

To test whether there was a direct effect on the catalytic capacity of CaI during long-term photocatalysis, the activity of CaI was monitored using the MV-dependent H<sub>2</sub> evolution assay. As shown in Figure 6b CaI alone showed no evidence of light-dependent inactivation (Figure 6b, green plot) compared to the enzyme kept in the dark (Figure 6b, blue plot), and both the light and dark incubated samples lost the same ~20% of activity during the 6 h period. When in a complex with CdS, CaI showed a similar level of stability as the free enzyme if incubated in the dark. However, when CdS:CaI complexes were illuminated, the CaI was clearly inactivated (Figure 6b, red plot). Notably, the inactivation kinetics closely followed the decline in H<sub>2</sub> production rate (Figure 6, part a vs b), with an acceleration in rate after 1 h. At the 4 h period, a small but significant (~20%) fraction of active CaI remained even after the photocatalytic rates had approached zero. Thus, the loss of catalytically active CaI results from a light-dependent process occurring within the MPA–CdS, but does not completely account for the loss in photocatalytic H<sub>2</sub> production by CdS:CaI.

It has been shown that long-term illumination of ligand-capped Cd chalcogenide nanocrystals results in the loss of surface thiol ligands.<sup>52,69,76</sup> On the basis of the effects of similar sulfur compounds on the activities of other metalloenzymes, we hypothesize that this process in MPA–CdS results in release of free MPA that in turn inhibits CaI activity.<sup>77–79</sup> To test this, the effect of MPA on CaI H<sub>2</sub> evolution activity was measured over time (Table 2). MPA led to inactivation of CaI in a time-

**Table 2. Effect of [MPA] on CaI Inactivation**

[MPA] (nM)	MPA:CaI molar ratio <sup>a</sup>	I <sub>50</sub> (h) <sup>b</sup>
19	0.5	6
38	1	4
76	2	3
190	5	2
380	10	0.75
760	20	>0.1

<sup>a</sup>38 nM CaI. <sup>b</sup>The CaI activity was measured as H<sub>2</sub> evolution using sodium dithionite reduced MV (5 mM)

dependent manner, with nanomolar concentrations effectively reproducing the loss in CaI activity in CdS:CaI complexes. This amount of MPA is ~0.2% of the MPA coverage on CdS nanorods, so deactivation of CaI occurred before MPA oxidation was sufficient to contribute to precipitation of CdS. We also tested the effect of Cd<sup>2+</sup> on CaI activity and found no effect at 760 nM Cd<sup>2+</sup> (Figure S6, Supporting Information), which is equivalent to the MPA concentration that led to maximal inactivation rates of CaI (Table 2). Together the data support the hypothesis that prolonged illumination of the CdS:CaI results in the slow light-driven loss of MPA from the nanorod surface, which in turn leads to the progressive loss of CaI activity. The mechanism of CaI inactivation by MPA is not yet clear and may arise from active site inhibition,<sup>79</sup> extraction

of the distal iron–sulfur clusters,<sup>77,80</sup> or a combination of both. Interestingly, the loss of CaI activity does not appear to be affected by the presence of AA (Figure 6c). Thus, if the free form of MPA accumulates through oxidation by photoexcited holes, this process is competitive with, or may be dominant over, hole-scavenging by AA. Moreover, as MPA molecules are lost from the CdS surface, the resulting defect sites may act as electron sinks. In combination, these effects would explain the lack of photocatalytic production of H<sub>2</sub> in the absence of AA.

## CONCLUSIONS

In this study, we present an analysis of the photocatalytic behavior of CdS:CaI complexes. The QY for H<sub>2</sub> production was 20%, with TOF values [mol H<sub>2</sub> (mol CaI)<sup>-1</sup> s<sup>-1</sup>] of up to 380 under 405 nm light and 983 under high intensity (30 000 μE m<sup>-2</sup> s<sup>-1</sup>) white light (Table S3, Supporting Information), significantly higher than the current values for PSI hybrid systems.<sup>16,18,20,81</sup> The high TOF was a result of directed assembly, in which site-specific binding of the MPA–CdS to a positive patch on the CaI surface facilitates rapid ET from CdS to the CaI electron transport chain iron–sulfur clusters. This molecular orientation was confirmed by competition assays, where CdS acted as an inhibitor of Fd-driven H<sub>2</sub> production by CaI. The activity of complexes was manipulated by changing the contributions of the individual photocatalytic processes. The CdS:CaI molar ratios controlled the contribution of productive ET, with the 1:1 coverage CdS:CaI complexes having the highest TOF. Overall H<sub>2</sub> production rates are reduced by the mismatch of excitation rate and enzyme TOF at solar-flux levels, which we hypothesize contributes to BET from CaI to CdS (as a form of hole-scavenging). The concentration of AA was shown to control the contribution of external HT in regeneration of the CdS ground state. Long-term stability was affected by photodegradation of the MPA–CdS, limiting the TON to a value of 10<sup>6</sup> H<sub>2</sub> before cessation of photocatalysis. The photodegradation was at least in part due to the oxidative loss of MPA from the CdS surface (Figure 1d), which could be improved through stabilizing surface ligands or enhancing the HT reaction to limit ligand photo-oxidation. Long-term efficiencies in other photocatalytic systems also suffer from photodegradation of the light-absorbing components, but due to different processes. For example, the instability of CdTe:CaI biohybrids observed previously might be due to energetic mismatch of the CdTe valence band and MPA oxidation potentials (in contrast to CdS as shown in Figure 1d) resulting in photodegradation of the CdTe. In dye-sensitized Ru(bpy)–TiO<sub>2</sub>–[NiFeSe]–H<sub>2</sub>ase complexes,<sup>22,23</sup> photocatalytic activity was lost due to photodegradation of the Ru(bpy)–TiO<sub>2</sub> and not to any loss in H<sub>2</sub>ase activity. ZnSe/CdS–Pt complexes also lose activity but are reactivated by addition of fresh mercapto-carboxylic acid ligands.<sup>30</sup>

The light absorption of semiconductor nanocrystals is robust and saturates at very high light intensities ( $\gg E\text{ m}^{-2}\text{ s}^{-1}$ ).<sup>82</sup> Photosynthesis, on the other hand, saturates at  $\sim 400\ \mu\text{E m}^{-2}\text{ s}^{-1}$  (measured as the O<sub>2</sub> evolution rate) or  $\sim 16\%$  of the peak photosynthetic active radiation (PAR) solar flux of 2500 μE m<sup>-2</sup> s<sup>-1</sup>.<sup>83</sup> Saturation arises from several factors, including catalysis and intermolecular ET rates that are diffusion limited. By direct coupling of H<sub>2</sub>ase, or another redox active catalyst, to CdS as the absorber, with ligands that allow for near optimal orientations and fast ET, the upper limits of catalytic capacity can be explored. We expect that by addressing surface ligand limitations, nanocrystal–CaI biohybrids can sustain TOF's that

approach the  $k_{\text{cat}}$  of CaI, illustrating the capacity of the extraordinary light-harvesting potential of nanocrystals, and fast catalytic rates of enzymes, for engineering direct photochemical solar energy conversion systems.

## ASSOCIATED CONTENT

### Supporting Information

Expression and purification protocol for ferredoxin, CdS nanorod synthesis, CdS nanorod potential determination, time-dependence of CdS:CaI self-assembly, Poisson distributions of CdS:CaI molar complexes for various CdS and CaI mixtures, effect of headspace H<sub>2</sub> on long-term CdS:CaI photocatalytic rates, QY measurement data, kinetic analysis of [AA] on photochemical H<sub>2</sub> production, CaI TOF dependence on CdS:CaI ratio and white light intensity, UV–vis absorption spectra during long-term H<sub>2</sub> generation, TEM images, and distributions of lengths and diameters at several time points. This material is available free of charge via the Internet at <http://pubs.acs.org>.

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### Notes

The authors declare no competing financial interest.

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